

01/10/00  
jc595 U.S. PTO

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

jc690 U.S. PTO  
09/479877  
01/10/00

Patent Application of  
Wolf, et al

Filed: January 10, 2000

Title: TRANSFORMED BACTERIA PRODUCING CS6 ANTIGENS AS VACCINES

Hon. Commissioner of Patents  
and Trademarks  
Washington, D.C. 20231

New Patent Application

Sir:

Transmitted herewith is a patent application entitled  
"TRANSFORMED BACTERIA PRODUCING CS6 ANTIGENS AS VACCINES". The  
inventors are Marcia K. Wolf, Fred J. Cassels and Edgar Charles  
Boedeker.


Enclosed are 29 pages of specification (including the claims),  
7 claims (5 independent) and 1 page of drawings (informal).  
Applicants request, in accord with 37 CFR 1.53 (a) and (b), that a  
serial number and filing date be assigned to the application as of  
the date of receipt of the presented papers. An executed  
Declaration and the filing fee will be forwarded in due course.

Please send all correspondence to:

Glenna Hendricks, Esq.  
P.O. Box 2509  
Fairfax, VA 22031-2509

Please direct all telephone calls to Glenna Hendricks, (703)  
591-4470.

Respectfully submitted;

  
Glenna Hendricks, Reg. No. 32,535

Attachments

## APPLICATION FOR UNITED STATES PATENT

**Inventor:** Marcia K. Wolf

This application is a continuation of patent application USSN 08/788,145 08/788,145 filed January 24, 1997, which is a continuation-in-part of patent application USSN 243,482 filed May 13, 1994, which is now abandoned.

### Field of the Invention:

This invention is related to a CS6 antigen for use in vaccines to protect from pathological effects of enterotoxigenic E. coli.

### Background of the Inventions:

CS6 is a component of CFA/IV (colonization factor antigen IV), one of three CFAs commonly found on enterotoxigenic Escherichia coli (ETEC). A recent study showed CS6 on 31% of ETEC isolated from soldiers in the Middle East. Other CFAs and similar proteins found on the surface of ETEC function as adhesins to attach bacteria to intestinal epithelial cells. Attached bacteria can then deliver their toxin(s) to the target cells. It has never been proved that CS6 is an adhesin for human tissue (Knutton, S., M. M. McConnell, B. Rowe, and A. S. McNeish, "Adhesion and Ultrastructural Properties of Human Enterotoxigenic Escherichia coli Producing Colonization Factor Antigens III and IV", Infect.Immun. **57**:3364--3371 (1989)), but a study in rabbits indicated CS6 is a colonization factor.

The CS6 operon has much in common with fimbrial operons from E. coli, Salmonella, Yersinia, Klebsiella, Haemophilus, and Bordetella. All contain molecular chaperons and ushers and a

number of structural subunits. This area contains two sequences homologous to insertion sequences, but no complete insertion sequences.

The low GC content (34%) and codon usage that is characteristic of *E. coli* genes that are expressed at low levels suggest the CS6 genes may have originated in another species. GC content of 35-45% is characteristic of Gram positive bacteria such as *Staphylococcus*, *Streptococcus*, *Bacillus*, and *Lactobacillus*. Low GC content is common for virulence-associated genes of *E. coli*.

CS6 is unusual because it is expressed on bacteria grown on a variety of media, unlike other CFA's from ETEC that are only expressed on bacteria grown on CFA agar. This unusual regulation is not peculiar to strain E8775 because ETEC isolated in 1990 expressed CS6 when grown on L agar. Temperature regulation of CS6 expression is characteristic of other CFA's from ETEC and virulence genes in a variety of pathogenic bacteria.

Although CS6 has never been visualized by negative staining, electron microscopy using anti-CS6 sera and colloidal gold indicated that it is present on the surface of ETEC. The apparent major protein associated with CS6 is approximately 16 kDa which is in the range of molecular weights typical for subunits for fimbriae and fibrillae. CS6 from ETEC strain E10703 of serotype O167:H5 has been cloned (Willshaw, et al., FEMS Microbio. Let. **49**: 473-478 (1988)). Only 3 kb of DNA was necessary for expression of CS6. That is in contrast to fimbriae that typically require approximately 9 kb of DNA and include genes for subunits as well as proteins

for transport of subunits and synthesis and assembly on the bacterial surface.

Grewal teaches bacterial strains transformed with plasmids containing genes which encode CS6. However, that reference does not teach use of plasmids under the controls of a lac promotor and a CS6 promotor.

#### **Brief Description of the Drawings:**

Figure 1 shows the restriction sites and the location of the pertinent genes that make up the CS6 operon.

Figure 2 shows derivation of the clone containing the kanamycin resistance gene.

#### **Description of the Invention:**

It is the purpose of this invention to provide structural proteins which will act as antigens to stimulate protective antibodies against enterotoxigenic Escherichia coli. Particularly important are proteins having the antigenic properties of the proteins encoded by the cssA and cssB genes. Constructs may be prepared which encode either one or both of the proteins. However, both proteins would be needed to provide desirable protection. The CS6 operon includes four genes which we designate cssA, cssB, cssC, and cssD. cssA and cssB encode the structural proteins of CS6. The CS6 operon has much in common with fimbrial operons from E. coli, Salmonella, Yersinia, Klebsiella, Haemophilus, and Bordetella. All contain molecular chaperons and ushers and a number of structural subunits. In a preferred embodiment, plasmids containing all four genes are transformed into attenuated bacteria, which are

then given by mouth to prevent morbidity arising from infection with E. coli.

CS6 has two major subunits; protein sequencing data demonstrates that CS6A and CS6B are both present. The DNA sequence yields a mechanism for expression of similar amounts of the two proteins. The CS6 operon contains DNA immediately downstream of cssB which can form a stem-loop with a stem rich in G and C which commonly act as transcription terminators. Termination at this site yields a transcript with cssA and cssB such that CssA and CssB proteins would be translated in equal amounts. Fimbrial operons for Pap, K99, and K88 have stem loops immediately downstream of the genes for the major coding structural subunits. This has been offered as a mechanism for over expression of subunit genes relative to other genes in the operons. In the case of CS6, this would allow over expression of both CS6A and CS6B.

The occurrence of two major structural proteins is unusual because fimbriae have a single major subunit and a number of minor subunits. CS3, which has been designated fibrillar rather than fimbrial, is an exception to this generality because it has 2 subunits. CssD belongs to the family of molecular ushers located in the outer membrane that accept subunits from the chaperone and escorts them to the bacterial surface. Apparently the entire cssD gene is not necessary for CS6 expression since CS6 was detected from clones carrying pDEP5 which only contains the N-terminal one-third of cssD. Klemm and Christiansen found that mutations in the usher for Type 1 fimbriae reduced fimbriation but 10% of the

bacteria produced a few fimbriae (Mol.Gen.Genet. 220:334-338).

The CS6 proteins are produced in the transformed bacteria and are present on the exterior surface of the bacteria. These proteins give rise to immunological response in the host. For immunization, the bacteria may be given either dead or alive. When attenuated bacteria have been transformed, the bacteria can be given live in mildly basic carriers. Economical and readily available carriers include carbonated water which may be flavored. The administration of the transformed bacteria in carbonated beverages is particularly useful, since the means necessary for administration is widely available.

In a preferred embodiment, the products are produced under the control of a lac promotor from pUC19. In the preferred embodiment, a vector pM346 containing a kanamycin resistant gene makes it possible to provide products which are appropriate for use in humans.

The CS6 proteins may also be extracted from the supernatant of the culture containing the organisms which express the proteins. The proteins may then be administered orally. The proteins may be formulated by means known in the art, including microencapsulation, coated capsules and liposomes. The proteins may be lyophilized before formulation.

## **MATERIALS AND METHODS**

### Source of nucleic acid

The genes for CS6 expression were from enterotoxigenic Escherichia coli (ETEC) strain E8775 tox<sup>-</sup> of serotype O25:H42 which

was a gift from Alejandro Cravioto. E8775 tox<sup>-</sup> is a derivative of E. coli strain E8775 which was originally isolated from Bangladesh. DH5 $\alpha$  which was purchased from Bethesda Research Laboratories, Inc., Gaithersburg, MD. pUC19 was originally purchased from P-L Biochem.

The antibiotic resistance gene encodes resistance to kanamycin and was purchased from Pharmacia, Uppsala, Sweden (Kan<sup>R</sup> GenBlock<sup>®</sup>).

CS6 expression is regulated from its native promoter. That is demonstrated by retention of control by growth temperature and is consistent with the DNA sequence determined from the clone. A contribution of the lac promoter from pUC19 is undefined. The contribution of increased copy number of the plasmid is probably substantial.

The nucleotide sequence containing the coding region was determined to be as constructed containing the kanamycin resistance gene was as follows:

```
1  AAGCTTGTA  CCAGTTGATA  AAAATATATC  ACGCTGGGAA  TGACGTGATG
51  TATATACGGA  GCAGCTATGT  CGGAACAGAT  ATTTTCCTAT  CGGTATGCGT
101 TGTGAGTAAG  CGTAAAGCCA  ATGCTGTCTG  TAACTCCTGA  TCCTTGCAGA
151 CTAAATTAGA  GCTCCTTCTA  AATTAGACGG  ATGGATAAAC  CTACAGACTG
201 GCGCTCTGGG  TCTCGCCGGA  TATTTTCTAA  TGAATTTAAG  CTTCATATGG
251 TTGAACTGGC  TTCGAAACCA  AATGCCAATG  TCGCACAACT  GGCTCGGGAA
301 CATGGCGTTG  ATAACAACCT  GATTTTAAAA  TAGCTACGCC  TCTGGCAAAG
351 AGAAGGACGT  ATTTCTCGTA  GAATGCCTCC  AACTATTGTA  GGCCCTACAG
401 TACCACTGAG  GTAGCCTGAA  TTTAAAGCCG  AAGCGGTCAG  AACTGTTCTT
451 GGTGTGAACG  TAGCACTCAC  CAATAAAAGC  ATCAATACGG  TGCTCTGTTG
501 ACACATTACG  AATGTTATGT  ATACAATAAA  AATGATTATA  GCAATATTAA
```

551 TGGTGTTATA TGAAGAAAAC AATTGGTTTA ATTCTAATTC TTGCTTCATT  
 601 CGGCAGCCAT GCCAGAACAG AAATAGCGAC TAAAAACTTC CCAGTATCAA  
 651 CGACTATTTT AAAAAGTTTT TTTGCACCTG AACCACGAAT ACAGCCTTCT  
 701 TTTGGTGAAA ATGTTGGAAA GGAAGGAGCT TTATTATTTA GTGTGAACTT  
 751 AACTGTTCCCT GAAAATGTAT CCCAGGTAAC GGTCTACCCT GTTTATGATG  
 801 AAGATTATGG GTTAGGACGA CTAGTAAATA CCGCTGATGC TTCCCAATCA  
 851 ATAATCTACC AGATTGTTGA TGAGAAAGGG AAAAAAATGT TAAAAGATCA  
 901 TGGTGCAGAG GTTACACCTA ATCAACAAAT AACTTTTAAA GCGCTGAATT  
 951 ATACTAGCGG GGAAAAAAA ATATCTCCTG GAATATATAA CGATCAGGTT  
 1001 ATGGTTGGTT ACTATGTAAA CTAAATACTG GAAGTATGAT TATGTTGAAA  
 1051 AAAATTATTT CGGCTATTGC ATTAATTGCA GGAAGTTCCG GAGTGGTAAA  
 1101 TGCAGGAAAC TGGCAATATA AATCTCTGGA TGTAAATGTA AATATTGAGC  
 1151 AAAATTTTAT TCCAGATATT GATTCCGCTG TTCGTATAAT ACCTGTTAAT  
 1201 TACGATTCCG ACCCGAAACT GGATTCACAG TTATATACGG TTGAGATGAC  
 1251 GATCCCTGCA GGTGTAAGCG CAGTTAAAAT CGCACCAACA GATAGTCTGA  
 1301 CATCTTCTGG ACAGCAGATC GGAAAGCTGG TTAATGTAAA CAATCCAGAT  
 1351 CAAAATATGA ATTATTATAT CAGAAAGGAT TCTGGCGCTG GTAACTTTAT  
 1401 GGCAGGACAA AAAGGATCCT TTCCTGTCAA AGAGAATACG TCATACACAT  
 1451 TCTCAGCAAT TTATACTGGT GGCGAATACC CTAATAGCGG ATATTCGTCT  
 1501 GGTACTTATG CAGGAAATTT GACTGTATCA TTTTACAGCA ATTAAAAAAA  
 1551 GGCCGCATTA TTGCGGCCAT TGACGATACT GCTAGGCAAA AATATGAAAT  
 1601 CAAAGTTAAT TATACTATTG ACGTTAGTGC CATTTTCATC TTTTCAACA  
 1651 GGAAATAATT TTGAAATAAA TAAGACACGA GTAATTTACT CTGACAGCAC  
 1701 ACCATCAGTT CAAATATCAA ATAATAAAGC ATATCCTTTA ATTATTCAAA  
 1751 GCAATGTATG GGATGAAAGC AATAATAAAA ATCATGACTT TATAGCAACA  
 1801 CCACCGATTT TTAAAATGGA AAGTGAAAGT CGGAATATAA TAAAAATAAT



1851	TAAAACAACCT	ATTAATTTTGC	CGGACTCTCA	GGAAAGTATG	AGATGGTTAT
1901	GTATTGAATC	AATGCCACCA	ATAGAAAAAA	GTAATAAAAT	AAACAGAAAA
1951	GAAGGAAGGA	CAGACAGTAT	TAATATCAGC	ATTCGGGGGT	GCATTAAACT
2001	GATATATCGA	CCTGCCAGTG	TTCCGTCTCC	TGTTTTTAAAT	AATATAGTAG
2051	AAAAATTAAA	ATGGCATAAA	AATGGAAAGT	ATCTTGTATT	AAAAAATAAT
2101	ACACCCTATT	ACATTAGCTT	TTCTGAGGTT	TTTTTTTGATT	CAGATAAAGT
2151	AAACAATGCA	AAAGATATTT	TATATGTAAA	ACCATACTCA	GAGAAGAAAA
2201	TAGATATCAG	CAACAGAATA	ATAAAAAAAA	TCAAATGGGC	TATGATTGAT
2251	GATGCTGGCG	CAAAAACAAA	ACTTTATGAA	TCAATTTTAT	AAAAAATCTC
2301	ATTACAGTAT	ACAAAAACAT	CAGATTACAG	GCTTGCTTTT	TTTGCTATTT
2351	ATATATCCTT	TCTCAACCTC	ATATGGAAAT	GAACAATTTA	GTTTTGACTC
2401	ACGATTCCCTA	CCATCAGGTT	ATAATTACTC	TTTAAATAGT	AACTTACCTC
2451	CTGAAGGTGA	GTATCTGGTT	GATATTTATA	TTAACAAAAT	AAAAAAGGAG
2501	TCCGCGATTA	TTCTTTTTTA	TATAAAAGGA	AATAAACTTG	TACCATGTTT
2551	ATCAAAAGAA	AAAATTTTCAT	CTTTGGGTAT	CAACATTAAT	AATAACGACA
2601	ACACAGAGTG	TGTAGAAACA	AGTAAGGCAG	GTATTAGTAA	TATCAGCTTT
2651	GAGTTTAGCT	CTCTTCGTTT	GTTTATTGCT	GTACCGAAAA	ATCTTCTGTC
2701	TGAGATTGAT	AAAATATCAT	CAAAGGATAT	AGATAACGGG	ATTCATGCTT
2751	TATTTTTTTAA	TTATCAAGTA	AATACAAGGC	TAGCCAATAA	TAAAAATCGT
2801	TATGATTACA	TTTCTGTTTC	ACCAAATATA	AATTATTTTTT	CATGGCGGTT
2851	GCGTAATCTT	TTTGAATTTA	ACCAAAACAA	CGATGAAAAA	ACATGGGAAA
2901	GAAACTACAC	TTATCTAGAA	AAAAGTTTTT	ATGATAAAAA	GCTAAACTTA
2951	GTCGTTGGTG	AAAGTTATAC	GAATTCAAAT	GTTTATAATA	ACTACTCTTT
3001	TACTGGTATT	TCAGTTTCTA	CAGATACAGA	TATGTATACG	CCAAGTGAAA
3051	TCGATTATAC	ACCAGAAATT	CATGGAGTGG	CTGATTCAGA	CTCTCAGATT
3101	ATTGTCAGGC	AAGGCAACAC	CATTATCATT	AATGAAAGTG	TTCCAGCCGG



4451 TTTAAGCTTA CAACTAAAGA TGGAAAAACG CCCCCATTAG GAGCTATAGC  
 4501 CCATGAAAAA AATGGAAAAC AGATTAATAC GGGTATCGTT GACGATGATG  
 4551 GTATGCTTTA TATGTCTGGA TTATCAGGGA CAGGGATTAT TAATGTAACA  
 4601 TGGAATGGAA AAGTCTGTTC ATTTCTTTT TCAGAAAAAG ATATATCTAG  
 4651 CAAACAATTA TCTGTTGTAA ATAAACAATG TTAGGTAGTG CATCCAATTA  
 4701 GTAGAACATG TGTTTTTCGA TAAACGCTCC GATCTCTTTT TCGTGGATCT  
 4751 CAACTGAGCG TGAGAAGCAG ATTGTTTTAC GAGCCAACCG CTTAATGCGG  
 4801 GTGCGTAGCG TCAGATTATT ACGCTCAATG CGTTGGGTGA ATATTTTGCC  
 4851 GGTCAGATGC TTATTCTTCG GTACC

Sequence ID No. 1

#### B. Cell expression clone:

E. coli HB101 was purchased from the American Type Culture Collection, Rockville, Maryland. It is ATCC #33694 and batch #91-1. (*Escherichia coli* ATCC 33694)

Preceptrol [Reg TM] culture. D. Ish-Horowicz and J.F. Burke HB101 <--- H. Boyer. Genotype: F- leuB6 proA2 recA13 thi-1 ara-14 lacY1galK2 xyl-5 mtl-1 rpsL20 supE44 hsdS20 (r- B m- B at least thi-hsd from *Escherichia coli* B). Produces isoprene (Curr. Microbiol. 30:97-103, 1995). J. Mol. Biol. 41: 459-472, 1969; Methods Enzymol. 68: 245-267, 1979.) Growth Conditions: Medium 1065 37C.

The plasmid containing the CS6 genes, the pUC19 origin of replication, and the gene for kanamycin resistance was transferred into E. coli HB101 by transformation. Transformants were selected by growth on L agar supplemented with 0.04% Xgal with 50 µgm per ml kanamycin sulfate and/or 50 µgm per ml ampicillin.

One copy of the CS6 genes exists as an extrachromosomal

plasmid of high (500-700) copy number. The CS6 genes are present on a plasmid, not integrated into the chromosome. The plasmid has been isolated from the strain and examined by agarose gel electrophoresis.

Plasmid DNA from E8775 tox<sup>-</sup> was transferred to laboratory strain DH5 $\alpha$  as a cointegrate with F' lac<sub>ts</sub>::Tn5, a conjugative plasmid. Transfer of the F' lac<sub>ts</sub>::Tn5 plasmid was selected by antibiotic resistance to kanamycin and CS6 expression was detected by Western blot using polyclonal antisera specific for CS6. Plasmids were isolated and a cointegrate was identified based on the large size. A spontaneous derivative in which the F' lac<sub>ts</sub>::Tn5 was removed was obtained and named M56. M56 contains a 61-megadalton plasmid from E8775 tox<sup>-</sup> and expresses CS6. Plasmid DNA from M56 was isolated, partially digested with restriction enzyme HindIII, and ligated to pUC19 that had been digested with HindIII. The ligation mixture was transformed into DH5 $\alpha$  and plated onto L agar supplemented with ampicillin and X-gal. White (lac<sup>-</sup>) colonies were picked to CFA plates supplemented with ampicillin and tested for CS6 expression.

A stable clone named M233 with an insert of approximately 24 kb into the cloning site of pUC19 was obtained. It was a spontaneous deletion of a larger clone. Subclones were obtained by digestion with various enzymes and a subclone containing approximately 5 kb from the HindIII site to KpnI site was found that expressed CS6. This clone was designated M285. Expression of CS6 was verified by transferring plasmids into E. coli strain HB101 and

detecting CS6 expression. The cloned CS6 is expressed under the same conditions as CS6 from the native 61-megadalton plasmid: CS6 was detected in extracts from bacteria grown at 37°C on CFA agar, L agar or MacConkey agar. CS6 was not expressed on bacteria grown at 17°C.

Studies were performed to determine appropriate handling of strain M285 for reproducible expression of CS6. Growth temperature was found to be especially important.

As indicated above, the protein sequence of the N-terminus of CS6 was determined from strains E8775 and from M233, the large clone derived from E8775. The 16 kDa proteins recovered from heat, saline extracts, and ammonium sulfate precipitation of M233 yielded two amino acids at each position (except cycle 12) indicating that two proteins were present. From the strength of the two signals, a probable primary sequence and a probable secondary sequence call was made for each of fifteen cycles. Quantitative analysis of the peak areas indicated that the molar ratio of the primary sequence (CS6A) to secondary sequence (CS6B) was approximately 3:1. The presence of the same two proteins was evident from strain E8775 grown on CFA agar and on L agar.

The DNA sequence of the DNA inserted into pUC19 in clone M285 was determined. Wim Gaastra's group in the Netherlands independently determined the DNA sequence of CS6 genes from ETEC strain E10703. The DNA sequences are available from Genbank accession numbers U04846 and U04844, respectively. A stretch of DNA of 4,219 base pairs was 98% identical. The DNA sequences diverge abruptly

on both sides of the common region, defining the limits of the CS6 genes. Four open reading frames were detected within the common area. These were designated cssA, cssB, cssC, and cssD.

The four open reading frames are preceded by consensus sequences for binding RNA polymerase and ribosomes. The first open reading frame, cssA was identified as the gene for the CS6 structural protein designated as the primary protein based on the amino acid N-terminal sequence. The deduced molecular weight agrees with that previously determined from SDS PAGE. cssA includes a signal sequence that is typical for many exported proteins. Eleven of 136 residues differ between the deduced Cssa proteins from E8775 and from E10704.

cssB begins 17 bases downstream from cssA. There is a typical signal sequence. cssB was identified as the gene for the CS6 structural protein designated as the secondary protein based on the amino acid N-terminal sequence. Five of 146 residues differ between the deduced Cssa proteins from E8775 and from E10704.

A region of dyad symmetry is present 6 bases downstream from CssaB in both clones. The sequence is GGCCGCATTATTGCGGCC (**Sequence #2**) in E8775 ETEC and GGCCGCATTATTGATTGCGGCC (**Sequence #3**) in E10703. Underlined bases form the G-C rich stem. The calculated free energy value of these structures is -14.8 kcal. Such structures are often found in fimbrial operons after the genes encoding structural proteins.

cssC begins 48 bases downstream from cssB. It has a typical signal sequence. The deduced proteins from both clones have 212

residues with 7 differences. A search of protein databases indicated C<sub>ss</sub>C is homologous to chaperone proteins necessary for expression of a number of fimbriae. The structure of PapD, the chaperone protein for Pap fimbriae, has been solved by X-ray crystallography and regions important for conserving the structural domains have been identified. C<sub>ss</sub>C conforms to this consensus.

The c<sub>ss</sub>D gene begins 14 bases upstream of the end of c<sub>ss</sub>C. The protein from E8775 is truncated relative to the protein from E10703 and there are 28 differences between C<sub>ss</sub>D from E8775 and E10703. The deduced protein from c<sub>ss</sub>D is homologous to molecular ushers. Overall, C<sub>ss</sub>D and the other proteins are only around 30% identical and around 50% similar, but the nine proteins have areas of high homology dispersed throughout, especially the first 410 residues, and 4 cysteines (residues 91, 112, and two near the C-terminus) which are conserved in all ushers.

A region of dyad symmetry is present 347 base pairs into the C<sub>ss</sub>D gene in both clones. The calculated free energy value of these structures is -7.2 kcal.

The plasmid from strain M285 was transformed into E. coli HB101 purchased from ATCC. The resulting strain was named M295. Expression of CS6 from M295 was achieved from small-scale fermentation. For production for human use, it was desirable to add a gene for resistance to kanamycin as the selectable marker. To that end, a vector was constructed based on pUC19 but with a gene for kanamycin resistance in place of the gene for ampicillin resistance. The CS6 genes from the pUC19 clone were subcloned into the

new vector and transformed into E. coli HB101.

Vector pM323 was constructed as follows. The kanamycin resistance gene was purchased from Pharmacia, Uppsala, Sweden (Kan<sup>R</sup> GenBlock<sup>®</sup>) and inserted into a cloning vector by Dr. David Lanar at WRAIR. DNA including the gene was amplified by PCR using the plasmid from Dr. Lanar as template and primers flanking the multiple cloning site. A product of the desired size (1,580 bp) was obtained, but with much template present. To increase the purity of the 1,580 fragment, a second PCR reaction was performed, this time with a small amount of the first PCR reaction as template. This product was confirmed by agarose gel electrophoresis, then digested with restriction enzyme HincII to remove unwanted restriction enzyme recognition sites. This product was ligated to pUC19 digested with SspI. The ligation mix was transformed into E. coli DH5 $\alpha$  and plated on L agar plates supplemented with kanamycin and Xgal. Isolate M318 had the desired phenotype of resistance to kanamycin and ampicillin with lacZ' intact. The gene for ampicillin resistance was removed to make a smaller vector. This was achieved by designing and synthesizing 2 oligonucleotides to amplify just the portion of pM318 with the gene for kanamycin resistance, the lacZ' gene carrying the multiple cloning site, and the origin of replication. PCR was performed, the product ligated then transformed into E. coli DH5 $\alpha$  with selection on L agar plates supplemented with kanamycin and Xgal. Isolate M323 had the desired phenotype of resistance to kanamycin, sensitivity to ampicillin, and intact lacZ'. Restriction digest



patterns confirmed the plasmid was a derivative of pUC19 with the gene for kanamycin resistance in place of the gene for ampicillin resistance.

The CS6 genes were cloned into vector pM323 from pM285. pM323 and pM285 were digested with restriction enzyme SstI, ligated, and transformed into E. coli DH5 $\alpha$  with selection on L agar plates supplemented with kanamycin and Xgal. Isolate M334 was determined to express CS6. Plasmid analysis revealed M334 carried the CS6 genes and 2 copies of the vector. An attempt was made to remove one copy of vector and at the same time move the clone into HB101, the desired host strain for fermentation. Isolate M340 was determined to express CS6 and retained 2 copies of the vector. An isolated colony of M340 was shown to produce high amounts of CS6 and was saved as M346.

In another embodiment lacking the kanamycin resistance gene, clones from an ETEC strain of serotype O25:H42 were derived from E. coli E8775 which was originally isolated from samples from Bangladesh. E. coli M56, which contains a 61-megadalton plasmid from E8775 Tox<sup>-</sup> and expresses CS6 has been described. The host for cloning was E. coli DH5 $\alpha$  which was purchased from Bethesda Research Laboratories, Inc., Gaithersburg, MD. The host for plasmids used for production of heat, saline extracts was HB101 (EMBO J. 4:3887-3893 (1985)).

Clones from E8775 were routinely grown in L broth. Antibiotics were added to L broth supplemented with agar as follows. Ampicillin was added, when appropriate, at 50  $\mu$ g/ml. Chlorampheni-

col was used at 30 µg/ml. X-Gal (5-bromo-4-chloro-3-indolyl β-D-galactopyranoside, Sigma) was added at 0.004%. CFA plates were prepared as previously described (Infect.Immun. 57:164-173 (1989)).

Cloning CS6 from E8775. The 61-megadalton plasmid from E. coli M56 was partially digested with HindIII and ligated to pUC19 that had been digested with HindIII. The ligation mixture was transformed into E. coli DH5α and plated onto L agar plates supplemented with ampicillin and X-gal. White (lac<sup>-</sup>) colonies were picked to CFA plates supplemented with ampicillin and tested for CS6 expression using antisera as described below. Plasmids were purified as described (Infect.Immun. 57:164-173 (1989)). Restriction enzymes were used according to the manufacturer's directions.

Detection of CS6 Expression. CS6 expression by bacterial colonies was detected after transfer to nitrocellulose and treatment as described by Mierendorf (Methods Enzymol. 152:458-469 (1987)). Primary antisera was specific for CS6 and was raised in rabbits and absorbed as previously described (Infect.Immun. 57:164-173 (1989)), except that rabbits were inoculated intravenously with live bacteria suspended in normal saline. Secondary antibody was peroxidase-conjugated goat anti-rabbit IgG (Cappel Laboratories, Cochranville PA) and detection was by TMB Substrate (Kirkegaard & Perry Laboratories, Inc., Gaithersburg MD).

Positive identification of CS6 was by western blots of heat, saline extracts. Heat, saline extracts were prepared from bacteria grown on the indicated media as described (Infect.Immun. 27:657-666 (1980)). Proteins were separated on precast 16% Tricine sodium

dodecyl sulfate-polyacrylamide gels (SDS-PAGE, Novex Novel Experimental Technology, San Diego CA) and transferred to nitrocellulose. Blots were handled as described above for colony blots.

Determination of N-terminal sequence. Heat, saline extracts were obtained from E8775 or clones of E8775 grown on L agar or CFA. Partial purification of CS6 was obtained by ammonium sulfate precipitation, with extracts sequentially precipitated at 20%, 40%, then 60% saturation. Samples at 40% and 60% saturation were dialyzed against deionized water and loaded onto precast 16% Tricine SDS-PAGE (Novex, San Diego, CA). Proteins were blotted onto polyvinylidene difluoride (PVDF) membranes (Westrans, Schleicher & Schuell, Keene, NH), stained by Coomassie blue (Rapid Coomassie Stain, Diversified Biotech, Newton, MA) and bands of approximately 16 kDa were excised for automated gas-phase N-terminal sequencing analysis (Applied Biosystems Model 470A, Foster City, CA). Data were analyzed using the Model 610A Data Analysis Program, Version 1.2.1 (Applied Biosystems, Inc, Foster City, CA). These methods have been described in detail (Infect.Immun. 60:2174-2181 (1992)).

DNA sequencing. DNA sequencing of the clones derived from E8775 was performed using the Model 373A DNA sequencing system from Applied Biosystems, Inc, Foster City, CA. Reactions were performed using the dideoxy method with fluorescent dye-labeled terminators, double-stranded templates, oligonucleotide primers, and AmpliTaq DNA polymerase following the manufacturer's protocol. Appropriate oligonucleotide primers were synthesized using a Model 391 DNA

Synthesizer (Applied Biosystems, Inc, Foster City, CA). Plasmids were purified for use as templates by a slight modification of the alkaline lysis method and cesium chloride density gradient centrifugation described by Maniatis (Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor (1982)). Plasmids were recovered by dialysis followed by multiple ethanol precipitations to remove residual salt. Sequence analysis was performed using software developed by the University of Wisconsin Genetics Computer Group (Nucleic Acids Res. **12**:387-395 (1984)).

## RESULTS

CS6 genes cloned from ETEC strain E8775 into pUC19. A stable clone named M233 was obtained from a partial digest of the 61-megadalton plasmid from E. coli M56. It was a spontaneous deletion of a larger clone. The insert in M233 was approximately 24 kb. Subclones were obtained by digestion with various enzymes and a subclone containing 4.9 kb from the HindIII site to KpnI was found that expressed CS6. This clone was designated M285. Expression of CS6 was verified by transferring plasmids into E. coli HB101 and detecting CS6 in heat, saline extracts. The cloned CS6 is expressed under the same conditions as CS6 from the native 61-megadalton plasmid (Table 1). CS6 was detected in western blots of heat, saline extracts of bacteria grown on CFA, L agar or MacConkey agar. CS6 was not expressed on bacteria grown at 17°C.

**Table 1. Regulation of CS6 Expression**

Strain:	M287	M56	E8775	HB101
Plasmid:	pM285	native	native	none
Chromosome:	HB101	HB101	native	HB101
Media	-----	-----	-----	-----
CFA 37°C	+	+	+	-
CFA 17°C	-	-	-	-
L agar	+	+	+	-
MacConkey	+	+	+	-

N-terminal sequence of CS6. The protein sequence of the N-terminus of CS6 was determined from strains E8775 and from M233, the large clone derived from E8775. The 16 kDa proteins recovered from heat, saline extracts, and ammonium sulfate precipitation of M233 yielded two amino acids at each position (except cycle 12) indicating that two proteins were present. From the strength of the two signals, a probable primary sequence and a probable secondary sequence call was made for each of fifteen cycles. Quantitative analysis of the peak areas indicated that the molar ratio of the primary sequence (CS6A) to secondary sequence (CS6B) was approximately 3:1. The presence of the same two proteins was evident from strain E8775 grown on CFA agar and on L agar.

DNA sequence of CS6 operons. The sequences of DNA cloned from E8775 (in M285) were determined. They are available from Genbank accession number U04846. The DNA sequence, when compared with sequences from another strain, were found to diverge abruptly on

both sides of the common area. Four open reading frames were detected. These were designated cssA, cssB, cssC, and cssD for CS six.

The GC content of the DNA is 34% and the codon usage is in the range found for Escherichia coli genes that are expressed at low or very low levels as defined by Osawa et al (Prokaryotic Genetic Code. Experientia 46:1097-1106 (1990)).

Genes encoding CS6 structural genes. The four open reading frames are preceded by consensus sequences for binding RNA polymerase and ribosomes. DNA and deduced amino acid sequence of cssA, a CS6 structural protein. The DNA sequence of the entire operon is available from Genbank accession number U04844. The deduced amino acid sequence from E8775 is given. The arrow indicates the site of cleavage of the signal peptide. The protein sequence is associated with the sequence for the second construct:

```

-35          -10          RBS
TTGACACATTACGAATGTTATGTATACAATAAAAAATGATTATAGCAATATTAATGGTGTTAT
ATGAAGAAAACAATTGGTTTAATTCTAATTCTTGCTTCATTCGGCAGCCATGCCAGAACA
M K K T I G L I L I L A S F G S H A R T 2
GAAATAGCGACTAAAACTTCCCAGTATCAACGACTATTTCAAAAAGTTTTTTTGCACCT
E I A T K N F P V S T T I S K S F F A P 22
GAACCACGAATACAGCCTTCTTTTGGTGAAAATGTTGGAAAGGAAGGAGCTTTATTATTT
E P R I Q P S F G E N V G K E G A L L F 42
AGTGTGAACTTAACTGTTCTGAAAATGTATCCCAGGTAACGGTCTACCCTGTTTATGAT
S V N L T V P E N V S Q V T V Y P V Y D 62
GAAGATTATGGGTTAGGACGACTAGTAAATACCGCTGATGCTTCCCAATCAATAATCTAC
E D Y G L G R L V N T A D A S Q S I I Y 82
CAGATTGTTGATGAGAAAGGGAAAAAAATGTTAAAAGATCATGGTGCAGAGGTTACACCT
Q I V D E K G K K M L K D H G A E V T P 102
AATCAACAAATAACTTTTAAAGCGCTGAATTATACTAGCGGGGAAAAAAAATATCTCCT
N Q Q I T F K A L N Y T S G E K K I S P 122
GGAATATATAACGATCAGGTTATGGTTGGTTACTATGTAAACTAA (Seq. #4)
G I Y N D Q V M V G Y Y V N * (Seq. #5) 136

```

The first open reading frame, cssA was identified as the gene for the CS6 structural protein CS6A designated as the primary protein based on the amino acid N-terminal sequence. cssA includes a signal sequence that is typical for many exported proteins. The deduced Cssa protein from E8775 has 136 residues, as shown above and in Table 2. The molecular weight agrees with that previously determined from SDS PAGE. No homologous proteins were found by searching the protein databases, but conserved residues are present near the C-terminus and this is typical of fimbrial subunits that are carried across the periplasm by chaperons.

**Table 2. Characteristics of Proteins Deduced from CS6 Operons**

<u>Protein</u>	<u>Source</u>	<u>Number of Residues</u>	<u>Molecular Weight</u>	<u>Isoelectric Point</u>
CssA (CS6A)	E8775	136	15,058	5.27
CssB (CS6B)	E8775	146	15,877	4.40
CssC	E8775	212	24,551	10.24
CssD	E8775	802	90,393	9.97

cssB begins 17 bases downstream from cssA. There is a typical signal sequence. cssB was identified as the gene for the CS6 structural protein CS6B designated as the secondary protein based on the amino acid N-terminal sequence. The C-terminus matches the consensus typical of fimbrial subunits. The sequence from E8775 is given. The arrow indicates the site of cleavage of the signal peptide.

```

MLKKIISAIA LIAGTSGVVN A GNWQYKSLDV NVNIEQNFIP DIDSAVRIIP 30
                        ↑
VNYDSDPKLD SQLYTVEMTI PAGVSAVKIA PTDSLTSSTGQ QIGKLVNVNN 80
PDQNMNYYIR KDSGAGNFMA GQKGSFPVKE NTSYTFSAIY TGGEYPNSGY 130
SSGTYAGNLT VSFYSN 146 (Seq. #6)

```

A region of dyad symmetry is present 6 bases downstream from CssB in both clones. The sequence is GGCCGCATTATTGCGGCC (**Seq. #2**) in E8775 ETEC. Underlined bases form the GC rich stem.

Genes with homology to fimbrial accessory proteins. cssC begins 48 bases downstream from cssB. It has a typical signal sequence. The deduced proteins from both clones have 212 residues with 7 differences. A search of protein databases indicated CssC is homologous to chaperone proteins necessary for expression of



Pap, CS3, K88, K99, CS31A, S, and Type 1 fimbriae of E. coli and SEF14 of Salmonella enteritidis, F1 and pH6 antigen of Yersinia pestis, Type 3 of Klebsiella pneumoniae, Type b of Haemophilus influenzae, and filamentous heamagglutinin of Bordetella pertussis. The structure of PapD, the chaperone protein for Pap fimbriae, has been solved by X-ray crystallography and regions important for conserving the structural domains have been identified. C<sub>ss</sub>C conforms to the following consensus. Below is the deduced amino acid sequence of c<sub>ss</sub>C. The \* indicates conservative amino acid replacements. Dots are gaps necessary for aligning all sequences. Boxes indicate beta strands as defined for PapD. The designation of the beta strands for domain 1 (A1 through G1) and domain 2 (A2-G2) are given below each box.

```

*      * *R***      * *      *** ** *      *****
NNF  EINKTRVIYS  DSTP  SVQISNN  KAYP..  LIIQSNVWDES  NNKNH..D  FIATPPIFKM
      A1          B1          C1          D1

*      * ****      ** E  ** * * *  P*      * * *
ESES  RNIIKIIK  TTI..NLPDSQE  SMRWLCIESM  PPIEKST..KINRKEGRTDSINISI  110
      E1          F1

      *K****  P* *      *      * * N  *P**  *** *
RGCIKLIYR  PASVPSPVFNN.IVEK  LKWHK  NGKY  LVLKN  NTPYY  ISFSEVF  160
      G1          A2          B2          C2

*      * *      P  * *      *      *      *D G*
FDSDKV..NNAKD  ILYVK  PY  SEKKID  ISN..RIIKKI  KWAMI  DDAGAKT  KLYESIL
      D2          E2          F2          G2

```

(Seq. #7)

CssD begins 14 bases upstream of the end of cssC. When compared with a second sequence there are 28 differences between CssD from E8775 and the other sequence. The deduced protein from cssD is homologous to molecular ushers found in operons of Pap, CS3, K88, K99 and Type 1 fimbriae of E. coli and SEF14 of Salmonella enteritidis, F1 of Yersinia pestis, and Type 3 of Klebsiella pneumoniae. Overall, CssD and the other proteins are only around 30% identical and around 50% similar. Asterisks above the CssD sequences indicates amino acids conserved relative to molecular ushers.

```

          ***** * ** * **
MNQFYKKSHYSIQKHQITGLLFLFIYPFSTSYGNEQFSFDSRFLPSGYN 50

  * ** G Y *** *N * * * C** * *
YSLNSNLPPEGEYLVDIYINKIKKESAIIPFYIKGNKLVPCLSKEKISSL 100

G* * * *C** ** * ** * ** * ** * **
GININNNDNTECVETSKAGISNISFEFSSRLRFIAVPKNLLSEIDKISSK 150

  * G* ***** ** * * * * W** *
DIDNGIHALFFNYQVNTRLANNKNRYDYISVSPNINYFSWRLRNLFEFNQ 200

          ***** * * *G* *** *** * G* * **
NNDEKTWERNYTYLEKSFYDKKLNLVVGESYTNNSVYNNYSFTGISVSTD 250

M * * * **A * * * * V**G*F**
TDMYTPSEIDYTPEIHGVADSDSQIIVRQGNTIINESVPAGPFSFPITN 300

* * * * * * * * * * * * * * *
LMYTGGQLNVEITDIYGNKKQYTVNNSSLPVMRKAGLMVYNFISGKLTKK 350

  * * *G ***** * * * * G** *G G*
NSEDGDFFTQGDINYGTHYNSTLFGGYQFSKNYFNLSTGIGTDLGFSGAW 400

* * * * * * * * * * * * * * *
LLHVSRSNFKNKNNGYNINLQONTQLRPFNAGVNFYAYRKKRYVELSDIG 450

  ** * ***** * * * * Y*
WHGNLYNQLKNSFSLSLSKSLNKYGNFSLDYNKMKYWDNAYDSNSMSIRY 500

          * ***P* * * **
FFKFMRAMITTNCSLNKYQSYEKKDKRFSINISLPLTKDYGHISSNYSFS 550

```

```

      *      ** **      *      *      *      *
NANTGTATSSVGLNGSFFNDARLNWNIQQNRTRNNGYTDNTSYIATSYA 600

      * * * *      * G * * *      * * *      * * * *
SPYGVFTGSYSGSNKYSSQFYASGGIVLHSDGVAFTQKAGDTSALVRID 650

      *      * * *      G * *      * *      * * * *      * *      *
NISDIKIGNTPGVYTGYNFALIPHLQPFKKNTILINDKGIPDGITLANI 700

      * * * *      *      *      * * *      * *      * * * *      *
KKQVIPSRGAIVKVKFDAKKGNDILFKLTTKDGKTPPLGAIAHEKNGKQI 750

      * * * *      * *      * * *      C      *
NTGIVDDDDGMLYMSGLSGTGIINVTWNGKVCSPFSEKDISSKQLSVVNK 800

      C
QC      802      (Seq. #8)

```

But comparison with the protein from another strain, the sequence data shows the proteins have areas of high homology dispersed throughout, especially the first 410 residues. C<sub>ss</sub>D has 4 cysteines (residues 91, 112, and two near the C-terminus) which are conserved in all ushers.

A region of dyad symmetry is present 347 base pairs into the C<sub>ss</sub>D gene in both clones. The calculated free energy value of these structures is -7.2 kcal.

DNA flanking the CS6 genes. When compared with another strain, the DNA sequences of the two clones diverge immediately downstream of C<sub>ss</sub>D and 96 bases upstream of C<sub>ss</sub>A. The non-homologous flanking regions have homology with five distinct insertion sequences. The homologies include 3% to 32% of each insertion sequence but not entire insertion sequences. The homology of and Iso-IS1 in E8775 continues beyond the clones we have sequenced and may be a complete insertion sequences in the native plasmids.

It should be noted that minor variation in bases of the peptides does not destroy antigenicity. A protein having at least 60% homology with the CS6 A and B proteins identified herein having conservative substitution would be expected to have desirable properties.

As indicated previously, bacteria transformed with plasmids which express the CS6-A and CS6-B proteins may be administered by mouth. If the transformed bacteria are attenuated strains, they may be delivered live. It is also possible to administer killed bacteria. Carbonated beverages such as carbonated water are particularly useful as carriers which are inexpensive. When the bacteria are administered in a carrier wherein the pH is not over 7, an antacid may be given with the bacteria.

The CS6 A and CS6 B proteins may also be at least partially purified and administered by mouth by means usually used in the art to deliver antigens to the intestinal tract, including in protected forms such as liposomes, microcrystals, microdroplets, as microencapsulated formulations or as enterically coated capsules.

**What we claim is:**

1. A method of inducing, in a susceptible host, the production of antibodies against CS6 protein comprising administration of a composition of matter comprising a bacteria transformed with a plasmid which contains genes cssA and cssB, all of the cssC and DNA sequence of cssD which encodes at least 802 amino acids (at least 2406 base pairs), an origin of replication, a Lac promotor, and a kanamycin resistance gene wherein said bacteria expresses both CS6A and CS6B proteins.
2. A method of claim 1 wherein the bacteria are in a pharmaceutically acceptable carrier.
3. A method of claim 2 wherein the carrier is a carbonated liquid.
4. A protein containing the amino acid sequence  
R T E I A T K N F P V S T T I S K S F F A  
P E P R I Q P S F G E N V G K E G A L L F  
S V N L T V P E N V S Q V T V Y P V Y D E  
D Y G L G R L V N T A D A S Q S I I Y Q I  
V D E K G K K M L K D H G A E V T P N Q Q  
I T F K A L N Y T S G E K K I S P G I Y N  
D Q V M V G Y Y V N.

5. A composition of matter comprising a protein of claim 4 in a pharmaceutically acceptable carrier.

6. A protein containing the sequence:

GNWQYKSLDV NVNIEQNFIP DIDSAVRIIP VNYDSDPKLD SQLYTVEMTI  
PAGVSAVKIA PTDSLTSSTGQ QIGKLVNVNN PDQNMNYYIR KDSGAGNFMA  
GQKGSFPVKE NTSYTFSAIY TGGEYPNSGY SSGTYAGNLT VSFYSN .

7. A composition of matter comprising the sequence of claim 6 in a pharmaceutically acceptable carrier.

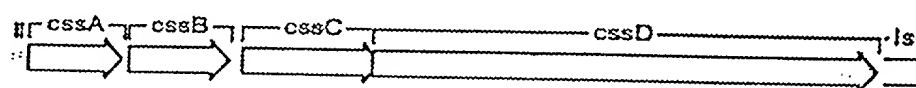
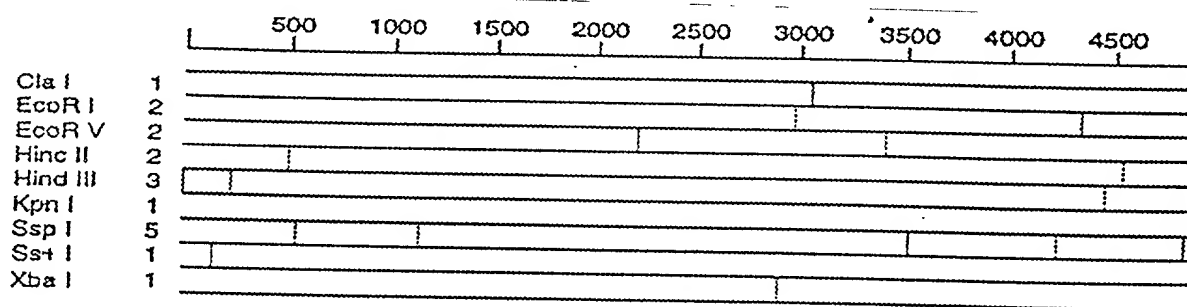


Fig 1

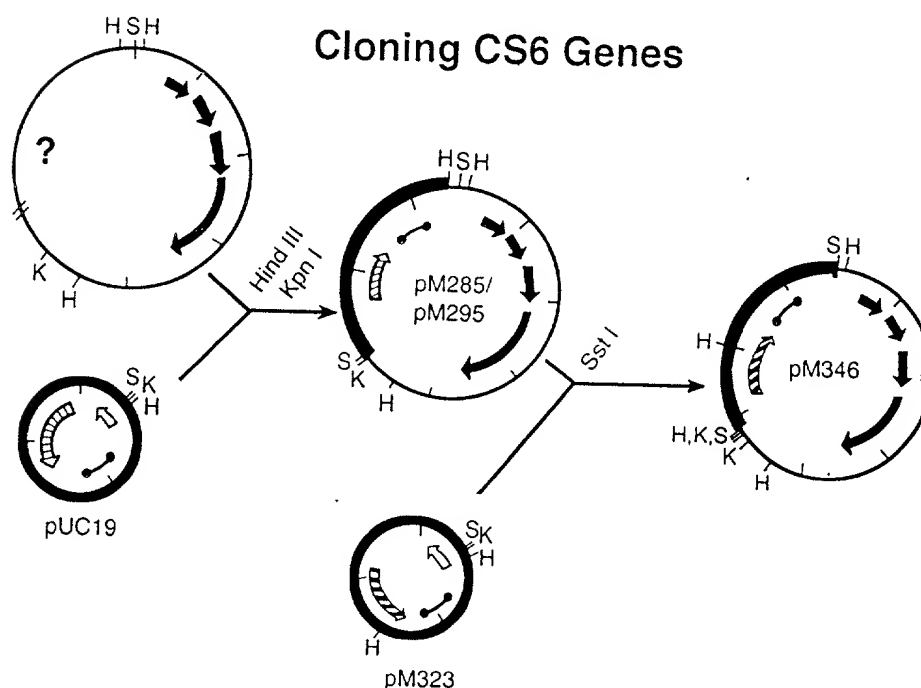


Fig. 2